

CONSTITUENTS FROM THE LEAVES OF *LANTANA CAMARA* (IV)

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Eight triterpenoids, betulonic acid (3), betulinic acid (4), oleanolic acid (5), lantadene A (6), lantadene B (7), icterogenin (8), lantanilic acid (9), and ursolic acid (10), three flavonoids, hispidulin (11), pectolarigenin (12), and pectolarin (13), as well as β -sitosterol-3-*O*- β -D-glucoside (2) and a mixture of campesterol (1a), stigmasterol (1b), and β -sitosterol (1c) were isolated from the leaves of the yellow flowering taxa of *Lantana camara* L. The structures of these compounds were established by spectroscopic methods and 2D NMR techniques.

Key words: *Lantana camara* L, Yellow flowering taxa, Triterpenoid, Flavonoid.

INTRODUCTION

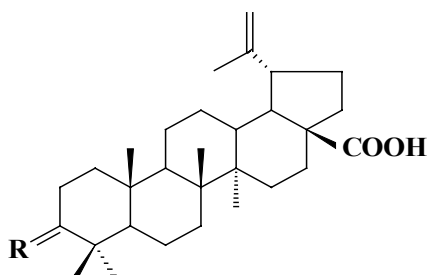
Lantana camara L. (Verbenaceae) is a rambling shrub with a number of flower colors viz. red, pink, white, yellow and violet¹. These plants are cultivated as ornamental or hedge plants in Taiwan².

Previously, we reported the phytochemical studies on the stems from the red, pink and yellow flowering taxa of *L. camara* L³⁻⁵. Herein, we report the isolation of thirteen compounds from the leaves of yellow flowering taxa of this plant, collected at Taichung, Taiwan.

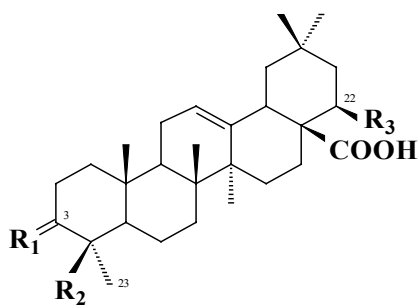
RESULTS AND DISCUSSION

Compounds **1-13** were isolated from the ethyl acetate extract of the leaves of this plant as described in the experimental section. Compound **1** was identified as a mixture of campesterol (**1a**), stigmasterol (**1b**) and β -sitosterol (**1c**) by spectroscopic method and GC analysis⁴. Compound **2** was identified as β -sitosterol-3-*O*- β -D-glucoside by spectroscopic and chemical methods⁴. In addition, betulonic acid (**3**)⁴, betulinic acid (**4**)³, oleanolic

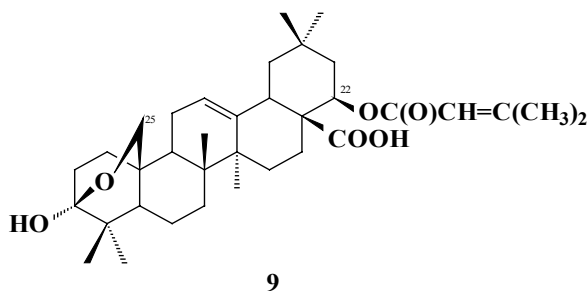
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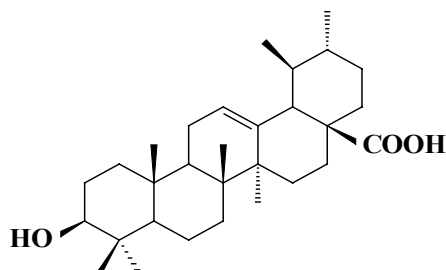
- 3** : R = O
4 : R = β -OH



- 5** : R₁ = β -OH R₂ = CH₃ R₃ = H
6 : R₁ = O R₂ = CH₃ R₃ = angeloyl
7 : R₁ = O R₂ = CH₃ R₃ = senecieryl
8 : R₁ = O R₂ = CH₂OH R₃ = angeloyl



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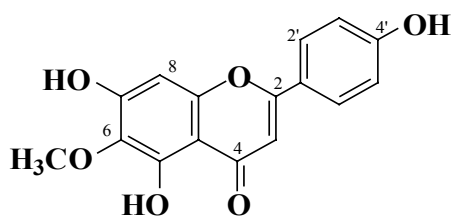


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acid (**5**)³, lantadene A (**6**)³, lantadene B (**7**)³, icterogenin (**8**)⁴, lantanilic acid (**9**)⁵, ursolic acid (**10**)³, and hispidulin (**11**)⁴ were also identified by comparing their spectral data with those in literature.

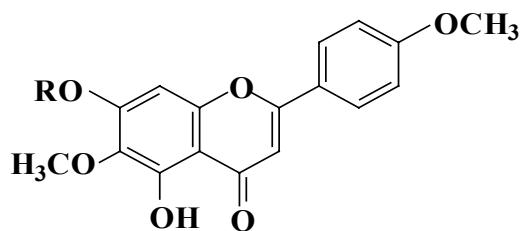
Compound **12**, obtained as yellow needles, mp: 210-211 °C, gave positive ferric chloride and Mg-HCl tests suggesting the presence of a flavonoid skeleton⁶. It showed an ion peak at m/z 315 ($[M + 1]^+$) which was 14 mass units more than hispidulin (**11**) in the FABMS spectrum. The UV spectral behavior of **12** with diagnostic reagents like NaOAc and AlCl₃ indicated the presence of the free 5- and 7-hydroxy groups⁷. Comparison of C-6 (δ 131.05) and C-8 (δ 94.05) chemical shifts in the ¹³C NMR spectrum of **12** with those (C-6, δ 131.11 and C-8, δ 94.05) of **11** showed the presence of 6-methoxy group⁴. The ¹H NMR spectrum showed an A₂X₂ system at δ 6.94 and 7.77 (each 2H, $J = 9.0$ Hz) due to a 4'-oxygenated B ring⁸. The attachment of methoxyl group to C-4' was confirmed by the correlation between H-3' and a methoxyl signal at δ 3.81 and between H-2' and H-3 in the NOESY spectrum. Furthermore, the HMBC spectrum indicated the correlations of H-3/C-2, C-4, C-1', H-8/C-6, C-7, C-9, C-10, and 6-OCH₃ (δ 3.86)/C-6. Thus, **12** was deduced as pectolarigenin and also confirmed by comparison of these data with those in the literature⁹.

Compound **13**, obtained as a yellow needle, mp > 300 °C, also gave positive ferric chloride and Mg-HCl tests. It possessed an ion peak at m/z 623 ($[M + 1]^+$) in FABMS spectrum. The NMR signals of **13** were similar to

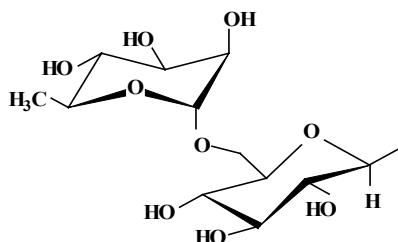


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12 : R = H



13 : R =



those of **12**. The only difference was the appearance of two additional glycosyl signals. Two anomeric protons [δ 5.26 (1H, *br s*, rhamnosyl H-1) and 5.63 (1H, *d*, $J = 7.3$ Hz, glucosyl H-1)] together with signals at δ 3.60 (1H, *dd*, $J = 11.5, 4.9$ Hz, glucosyl H_a-6), 3.75 (1H, *dd*, $J = 11.5, 2.5$ Hz, glucosyl H_b-6) and 1.18 (3H, *d*, $J = 6.3$ Hz, rhamnosyl H-6) revealed the rutosyl moiety. The position of rutosyl moiety on C-7 was confirmed by the HMBC correlation between glucosyl H-1 (δ 5.63) and C-7 (δ 157.7). Acid hydrolysis of **13** yielded an aglycone pectolinarigenin (**12**), D-glucose and L-rhamnose which were identified by the ¹H NMR and thin-layer chromatography. According to the coupling constant, the configuration of D-glucose and L-rhamnose was determined as β - and α -linkage, respectively¹⁰. The ¹³C NMR data of **13** also supported the attachment of rutosyl moiety to the 7-position of pectolinarigenin. Thus, **13** was identified as pectolinarin and also confirmed by comparison these data with those in the literature¹¹. This is the first report of the isolation of **12** and **13** from the genus *Lantana*.

EXPERIMENTAL

Melting points were measured on a Yanaco micro melting point apparatus and are uncorrected. IR spectra were recorded on a HITACHI I-2001 infrared spectrometer. EIMS and FABMS spectra were measured by Finnigan MAT 95 XL spectrometer. NMR spectra were recorded on Varian VXR-300 or Unity Inova 600 MHz spectrometers with TMS as an internal standard. Optical rotations were measured on a Perkin Elmer 241 MC digital polarimeter. UV spectra were recorded on a Shimadzu UV 260 ultraviolet spectrometer.

Plant Material

The leaves of *Lantana camara* L. (yellow flower) were collected in the campus of Providence University, Taichung in July 2000. The plant was identified by Mr. Nien-Yung Chiu (Institute of Chinese Pharmaceutical Sciences, China Medical University).

Extraction and Separation

The air-dried leaves (11.9 Kg) of *L. camara* were extracted with MeOH under reflux. The extract was concentrated *in vacuo*. The residue (660 g) was suspended in water and consecutively extracted with ethyl acetate and *n*-butanol. The ethyl acetate fraction (185 g) was chromatographed on a silica gel column using mixtures of CHCl₃-MeOH as eluent to give three fractions. Fraction I was purified by repeated column chromatography on silica gel eluting with *n*-hexane-ethyl acetate in order of increasing polarity to afford phytosterols (**1**, 25 mg), betulonic acid (**3**, 70 mg), betulinic acid (**4**, 10 mg), oleanolic acid (**5**, 1510 mg) and ursolic acid (**10**, 15 mg). Fraction II was further chromatographed on a silica gel column with *n*-hexane by increasing amounts of acetone to yield lantadene A (**6**, 20 mg), lantadene B (**7**, 210 mg), icterogenin (**8**, 25 mg) and lantanilic acid (**9**, 5120 mg). Fraction III was rechromatographed on a silica gel column with chloroform and methanol as eluent to give hispidulin (**11**, 780 mg), pectolarigenin (**12**, 5 mg), pectolarin (**13**, 25 mg) and β -sitosteryl-3-*O*- β -D-glucoside (**2**, 10 mg).

Pectolarigenin (12)

Yellow needles (CHCl₃/MeOH), mp: 210-211 °C; IR (KBr) ν_{\max} cm⁻¹: 3320 (OH), 1660, 1580; UV λ_{\max} nm (MeOH) (log ϵ): 276 (4.10), 333 (4.20), λ_{\max} nm (MeOH + NaOMe) (log ϵ): 276 (4.22), 296 (4.12), 368 (4.02); λ_{\max} nm (MeOH + NaOAc) (log ϵ): 276 (4.27), 297 (4.10), 368 (4.04); λ_{\max} nm (MeOH + AlCl₃) (log ϵ): 283 (4.06), 300 (4.11), 354 (4.23); ¹H-NMR (600 MHz, CDCl₃/CD₃OD): δ 3.81 (3H, *s*, 4'-OCH₃), 3.86 (3H, *s*, 6-OCH₃), 6.47 (1H, *s*, H-3), 6.48 (1H, *s*, H-8), 6.94 (2H, *d*, *J* = 9.0 Hz, H-3' and H-5'), 7.77 (2H, *d*, *J* = 9.0 Hz, H-2' and H-6'), 13.08 (1H, *br s*, 5-OH); ¹³C-NMR (150 MHz, CDCl₃/CD₃OD): δ 60.5 (*q*, 6-OCH₃), 55.3 (*q*, 4'-OCH₃), 164.2 (*s*, C-2), 103.3 (*d*, C-3), 182.8 (*s*, C-4), 153.1 (*s*, C-5), 131.1 (*s*, C-6), 156.4 (*s*, C-7), 94.1 (*d*, C-8), 152.3 (*s*, C-9), 105.0 (*s*, C-10), 123.3 (*s*, C-1'), 127.9 (*d*, C-2' and C-6'), 114.3 (*d*, C-3' and C-5'), 162.5 (*s*, C-4'); FABMS *m/z* (rel. int.%): 315 [M + 1]⁺ (73), 289 (26), 217 (10), 154 (100), 136 (74).

Pectolarin (13)

Yellow needles (CHCl₃/MeOH); mp > 300 °C; IR (KBr) ν_{\max} cm⁻¹: 3390 (OH), 1660, 1610; UV λ_{\max} nm (MeOH) (log ϵ): 276 (4.10), 333 (4.20), λ_{\max} nm (MeOH + NaOMe) (log ϵ): 276 (4.22), 296 (4.12), 368 (4.02); λ_{\max} nm (MeOH + NaOAc) (log ϵ): 276 (4.27), 297 (4.10), 368 (4.04); λ_{\max} nm (MeOH + AlCl₃) (log ϵ): 283

(4.06), 300 (4.11), 354 (4.23); ¹H-NMR (600 MHz, C₅D₅N): δ 1.18 (3H, *d*, *J* = 6.3 Hz, H-6, Rha), 3.60 (1H, *dd*, *J* = 11.5, 4.9 Hz, H_a-6, Glc), 3.70 (3H, *s*, 4'-OCH₃), 3.75 (1H, *dd*, *J* = 11.5, 2.5 Hz, H_b-6, Glc), 4.04 (3H, *s*, 6-OCH₃), 5.26 (1H, *s*, H-1, Rha), 5.63 (1H, *d*, *J* = 7.3 Hz, H-1, Glc), 6.88 (1H, *s*, H-3), 7.26 (2H, *d*, *J* = 9.0 Hz, H-3' and H-5'), 7.32 (1H, *s*, H-8), 8.05 (2H, *d*, *J* = 9.0 Hz, H-2' and H-6'), 13.60 (1H, *s*, 5-OH); ¹³C-NMR (150 MHz, C₅D₅N): δ 55.4 (*q*, 4'-OCH₃), 60.8 (*q*, 6-OCH₃), 164.7 (*s*, C-2), 104.2 (*d*, C-3), 183.1 (*s*, C-4), 154.0 (*s*, C-5), 133.8 (*s*, C-6), 157.7 (*s*, C-7), 95.0 (*d*, C-8), 153.1 (*s*, C-9), 107.1 (*s*, C-10), 123.5 (*s*, C-1'), 128.8 (*d*, C-2' and C-6'), 115.2 (*d*, C-3' and C-5'), 163.0 (*s*, C-4'), 102.3 (*d*, C-1, Glc), 74.6 (*d*, C-2, Glc), 78.4 (*d*, C-3, Glc), 71.2 (*d*, C-4, Glc), 77.6 (*d*, C-5, Glc), 67.5 (*t*, C-6, Glc), 102.4 (*d*, C-1, Rha), 72.0 (*d*, C-2, Rha), 72.8 (*d*, C-3, Rha), 74.0 (*d*, C-4, Rha), 69.8 (*d*, C-5, Rha), 18.5 (*q*, C-6, Rha); FABMS *m/z* (rel. int.%): 623 [M + 1]⁺ (2), 460 (3), 307 (27), 289 (17), 154 (100).

Acid hydrolysis of 13

A solution of **13** (10 mg) in 5% H₂SO₄ (10 mL) was refluxed for 4 hr. The solid, separated after cooling, was recrystallized from MeOH to give yellow needles (3 mg) of an aglycone (**12**), mp 210-211 °C. The aqueous layer was concentrated *in vacuo*. D-glucose and L-rhamnose were identified by TLC on cellulose in pyridine-EtOAc-HOAc-H₂O (5:5:1:3) R_f 0.4 and 0.6, respectively.

ACKNOWLEDGMENTS

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黃花馬纓丹葉部成分之研究

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本研究由黃花馬纓丹 [*Lantana camara* Linn. (yellow taxa)] 葉部分離得到十三個成分，經光譜分析確定其結構分別為 phytosterols (1), β -sitosterol-3-O- β -D-glucoside (2), betulonic acid (3), betulinic acid (4), oleanolic acid (5), lantadene A (6), lantadene B (7), icterogenin (8), lantanilic acid (9), and ursolic acid (10), hispidulin (11), pectolarigenin (12)及 pectolarin (13)。其中 pectolarigenin (12)及 pectolarin (13)是馬纓丹屬植物文獻上未曾報導的成分。

關鍵詞：黃花馬纓丹，馬鞭草科，三萜類，黃酮類，黃酮類配醣體。